



## Pb (II) sorption by acidically modified *Cicer arietinum* biomass

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### ABSTRACT

In this study, efficacy of native and acidically modified *Cicer arietinum* pod biomass was investigated for Pb (II) removal using metal ion concentration, kinetic, sorbent dose, pH and pretreatment as operational parameters. The metal uptake of biomass increased as a result of pretreatments and maximum sorption capacity (171.28 mg/g) was shown by H<sub>3</sub>PO<sub>4</sub> modified biomass. The sorption capacity increased in the order H<sub>3</sub>PO<sub>4</sub> > H<sub>2</sub>SO<sub>4</sub> > HCl > native, with increase in metal concentration. The equilibrium data was analysed using different sorption models to know sorption mechanism and results of ANOVA showed that Dubinin–Radushkevich model is fitted well to the experimental data. Sorption rate was slow because equilibrium was attained within 7 h and rate was best described by pseudo-first-order, second-order rate equation, Bangham and intraparticle diffusion models. Sorption capacity was maximum at pH 4 and it was observed also that sorption capacity decreased but % removal increased with increase in biosorbent dose.

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### 1. Introduction

The current pattern of industrial activity alters the natural flow of materials and introduces novel chemicals into the environment [1]. The rate at which effluents are discharged into the environment especially water bodies is increasing as a result of urbanization. Most of these effluents contain toxic substances especially heavy metals. The presence of heavy metals in the environment is of major concern because of their toxicity, bio-accumulating tendency, threat to human life and the environment [2,3]. Lead is widely used in many industrial applications such as storage manufacturing battery, printing, pigments, fuels, photographic materials and explosive manufacturing [4]. Lead is highly toxic as its presence in drinking water above the permissible limit (5 ng/mL) causes adverse health effects such as anemia, encephalopathy, hepatitis and nephritic syndrome [5].

Conventionally, the methods employed for the removal of heavy metals from effluents include oxidation and reduction, precipitation, filtration, electrochemical treatment and evaporation [6]. These physiochemical methods have several disadvantages such as unpredictable metal ion removal, high reagent requirements and formation of sludge and its disposal, in addition to high installation and operational costs [7]. Biosorption, a biological method of

environment control, can be an alternative to conventional waste-treatment facilities. Several workers have reported on the potential use of agricultural by-products as good substances for the removal of metal ions from aqueous solutions and wastewaters [8]. The ability of some agricultural by-products to adsorb heavy metals from wastewater and aqueous metal solutions has been reported in literature and these include cassava waste [9], sunflower stalk [10], waste tea [11], distillery sludge of sugar-cane industry [12], wheat bran [13] and horseradish tree [14]. Also, research by Park et al. [15] showed that chemical modification of agricultural adsorbents increased the sorption capacity of the adsorbent, thereby increasing the efficiency of the adsorbent.

In this study, pods of *Cicer arietinum* were chosen as a biosorbent for the removal of lead (II) from aqueous solutions because of the composition of *C. arietinum* [16] and its availability. *C. arietinum* belongs to family Leguminosae. Pakistan contributes up to 4.64% of the total *C. arietinum* production in the world. In Pakistan, 933,900 ha area was used for cultivation of *C. arietinum* in 2002 and production was 362,100 tonnes. Now, cultivation of *C. arietinum* has increased in Pakistan. Since removal of heavy metals using *C. arietinum* is already reported [17], biosorbent was chemically modified to observe enhancement or decrease in biosorption capacity of biomass in this study.

### 2. Material and methods

#### 2.1. Materials

Pods of chickpea (*C. arietinum*) were collected from a field near Chakwal, Pakistan. All pods were washed with distilled water in

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order to remove all particulate material. Then these pods were dried at room temperature and ground using food processor (Moulinex, France). The ground material was divided into four equal parts. Three parts were taken and treated with HCl, H<sub>2</sub>SO<sub>4</sub> and H<sub>3</sub>PO<sub>4</sub> (Fluka chemicals) in the ratio of 5 g sorbent/100 mL of 1 M reagent, respectively using orbital shaker (Gallenkamp) for 24 h. These acidically modified biomasses were filtered and washed with distilled water up to neutral pH. These washed pretreated biomasses were dried at room temperature and ground using food processor and then sieved (63 and 150 μm) through Octagon sieveer (OCT-DIGITAL 4527-01) and stored in airtight plastic containers. The fourth untreated biomass part was also sieved and used as native biomass.

Stock Pb (II) solution (1000 mg/L) was prepared by dissolving 1.598 g of Pb(NO<sub>3</sub>)<sub>2</sub> (Fluka chemical) in some distilled water and making volume up to 1000 mL using distilled water. Pb (II) solutions of different concentrations (50–200 mg/L) were prepared by adequate dilution of the stock solution with distilled water.

## 2.2. Batch sorption studies

In all experiments, 100 mL of Pb (II) solution (100 mg/L) having pH 4 was mixed with biosorbent (0.05 g) having a definite mesh size at 30 °C and 200 rpm in orbital shaker. To evaluate the effect of initial metal concentration, 100 mL solution having concentration (50–200 mg/L) of Pb (II) was stirred with native and pretreated biosorbents (0.05 g) having mesh size (63 and 150 μm) for 24 h. Further, the effect of contact time (5–480 min), effect of biosorbent dose (0.05, 0.1, 0.2 g) and effect of pH (4, 7 and 9) with 150 μm sieved H<sub>3</sub>PO<sub>4</sub> modified biomass were also evaluated. To adjust pH of solution, 0.1N HCl and NaOH were used. At the end of experiments, flasks were removed from shaker and solutions were filtered. Filtrates were stored in pre-cleaned and acid (HNO<sub>3</sub>) washed airtight plastic containers and analysed by PerkinElmer AAnalyst atomic absorption spectrophotometer equipped with lead hollow cathode lamp operating at wavelength of 232 nm and slit as 0.2 nm. Blank experiments were also carried out.

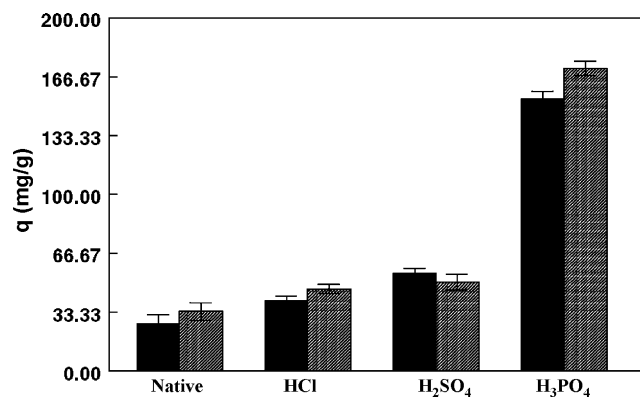


Fig. 2. Effect of pretreatment and particle size on biosorption of Pb<sup>2+</sup> using 63 μm ■ and 150 μm ▨ sieved native, HCl, H<sub>2</sub>SO<sub>4</sub> and H<sub>3</sub>PO<sub>4</sub> modified biomasses.

## 2.3. Calculations

The sorption capacity 'q' (mg/g) was calculated by simple concentration difference method:

$$q = \frac{V(C_i - C_e)}{M} \quad (1)$$

where C<sub>i</sub> and C<sub>e</sub> (mg/L) are initial and final metal concentrations, V is volume of the solution (L) and M is mass of biosorbent (g). Each experiment was conducted in triplicate and data represents mean of three independent experiments. Two-way analysis of variance (ANOVA) was done to evaluate the concentration data. All calculations were made using Slide Write version 3.0 and Microsoft Office 2000.

## 3. Results and discussion

### 3.1. Effect of metal concentration

The amount of metal ion sorbed per unit mass of native and pretreated biosorbents (i.e., sorption capacity) was found to increase

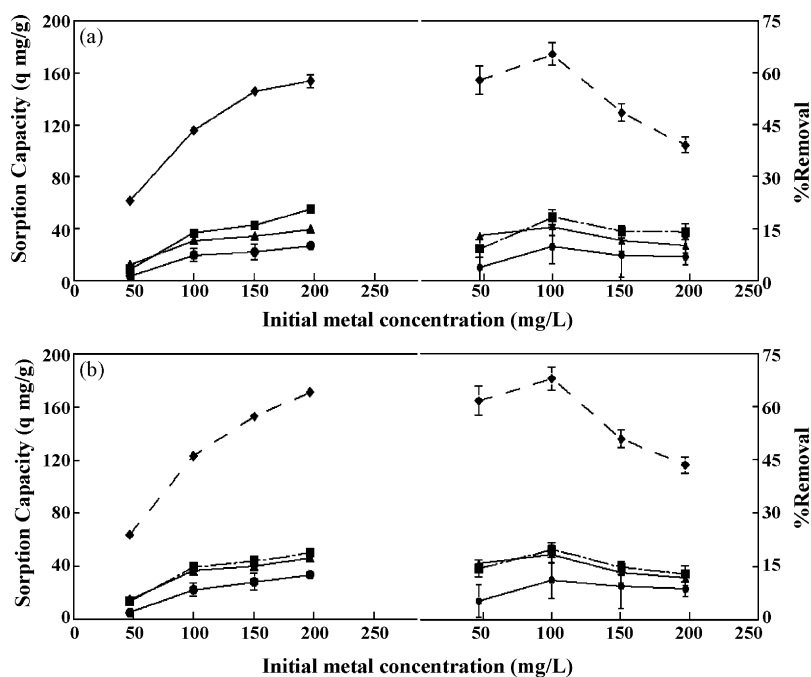


Fig. 1. (a) Effect of initial metal concentration on the biosorption of Pb<sup>2+</sup> using 63 μm sieved native ●, HCl ▲, H<sub>2</sub>SO<sub>4</sub> ■ and H<sub>3</sub>PO<sub>4</sub> ◆ modified biomasses. (b) Effect of initial metal concentration on the biosorption of Pb<sup>2+</sup> using 150 μm sieved native ●, HCl ▲, H<sub>2</sub>SO<sub>4</sub> ■ and H<sub>3</sub>PO<sub>4</sub> ◆ modified biomasses.

with increase in concentration of metal ion whereas percentage removal was maximum at 100 mg/L for all biomasses and then found to decrease with increase in initial metal ion concentration (Fig. 1a and b). The increase in sorption capacity may be explained by the fact that at low concentrations, the available metal occupies adsorption sites more quickly. However, at higher concentrations, metals diffuse to the biomass surface by intraparticle diffusion. The decrease in percentage removal is explained by the fact that at lower concentrations, almost all the ions were adsorbed very quickly and further increase in initial metal ion concentration led to saturation of biomass surface [18].

### 3.2. Effect of pretreatment and biosorbent size

Comparing the results, one can notice that the pretreatment of biomass by HCl, H<sub>2</sub>SO<sub>4</sub> and H<sub>3</sub>PO<sub>4</sub> (Fig. 2) increased the sorption capacity (mg/g) of biomass in order H<sub>3</sub>PO<sub>4</sub> (171.28) > H<sub>2</sub>SO<sub>4</sub> (50.4) > HCl (46.3) > native (33.62) for 150 μm sieved biomass as compared to 63 μm sieved H<sub>3</sub>PO<sub>4</sub> (153.8) > H<sub>2</sub>SO<sub>4</sub> (55.08) > HCl (39.7) > native (26.9). Generally, acid-treatment has been used for cleaning the cell wall and replacing the natural mix of ionic species bound on the cell wall with protons and other functional groups exposing new active sites for metal removal causing increase in sorption capacity [19–21]. Park et al. [22] also reported an increase in sorption capacity of biomass as a result of acidic pretreatment of *Ecklonia* biomass. H<sub>3</sub>PO<sub>4</sub> treatment showed remarkable increase in sorption capacity of biomass but a slight difference was observed in case of H<sub>2</sub>SO<sub>4</sub> and HCl treatments when compared to native biomass. Pretreatment of biomass with acids, apart from removal of mineral matter also resulted in introduction of oxygen surface complexes that change the surface chemistry by increasing the porosity and surface area of original sample. H<sub>3</sub>PO<sub>4</sub> pretreated biomass showed more sorption capacity and it was due to solubility of more mineral matter of biomass which resulted more porosity in biomass due to increased cellular mass [12]. When compared results of particle size (63 and 150 μm), 150 μm sieved biomass showed more sorption capacity as compared to 63 μm sieved biomass (Fig. 2). This behavior was also reported by Leusch et al. [23] who showed that larger biomass particles of *Sargassum fluitans* and *Ascophyllum nodosum* had higher metal uptake than smaller particles in the case of cadmium, copper, nickel, lead and zinc.

### 3.3. Equilibrium studies

In order to optimize the design of a sorption system for the removal of metals from wastewater, it is important to establish the most appropriate correlation for equilibrium curves. Experimental data acquired at different conditions is fitted to different models of sorption isotherms because equilibrium isotherm models are used to describe the experimental data. The equation parameters and the thermodynamics of these equilibrium models often provide some insight into the sorption mechanisms, surface properties and affinities of the sorbent (Fig. 3a and b). There are many theories relating to adsorption equilibrium.

#### 3.3.1. Freundlich isotherm model

The Freundlich isotherm [24] is originally empirical in nature but was later interpreted as sorption to heterogeneous surface or surface supporting sites of varied affinities. It is assumed that the stronger binding sites are occupied first and the binding strength decreases with the increasing degree of site occupation. Freundlich model is expressed as

$$\log q_e = \frac{1}{n} \log C_e + \log k \quad (2)$$

where  $1/n$  and  $k$  are Freundlich constants. The Freundlich isotherm parameter  $1/n$  measures the adsorption intensity of metal ions on biomass and Freundlich constant  $k$  is adsorption capacity of biomass. The value of  $1/n$  greater than unity indicates that adsorption can take place even at high concentrations but values of  $1/n$  in case of native biomass oppose the experimental data. The high  $1/n$  value of native biomass in comparison to acidically modified biomass indicated the sorption ability of native biomass but experimental data showed that pretreatment causes enhancement in sorption capacity of biomass. The value of  $k$  (dm<sup>3</sup>/g) is greater for H<sub>3</sub>PO<sub>4</sub> modified biomass as compared to other biomasses in favor of experimental data but order of  $k$  (dm<sup>3</sup>/g) values is disturbed in case of other biomasses. Values of  $R^2$  in case of H<sub>3</sub>PO<sub>4</sub> modified biomass are 0.9773 and 0.9711 for 150 and 63 μm sizes that are equal but greater than  $R^2$  values of native, HCl and H<sub>2</sub>SO<sub>4</sub> as indicated from Table 1. Freundlich isotherm model favorably satisfies the experimental data only in case of H<sub>3</sub>PO<sub>4</sub> modified biomasses.

#### 3.3.2. Langmuir isotherm model

The Langmuir isotherm theory assumes monolayer coverage of adsorbate over a homogeneous adsorbent surface [25]. Langmuir isotherm can be characterized by a plateau graphically. A saturation point reaches at equilibrium where no further adsorption occurs. Adsorption is assumed to take place at specific homogeneous sites within the adsorbent. Once a sorbate molecule occupies a site, no further adsorption takes place at that site. Linearized form of Langmuir model is:

$$\frac{C_e}{q_e} = \frac{1}{X_m K_L} + \frac{C_e}{X_m} \quad (3)$$

where  $K_L$  (dm<sup>3</sup>/g) is Langmuir constant termed as apparent energy of adsorption and  $X_m$  (mg/g) is sorption capacity. When values of  $R^2$  were compared, sorption phenomena did not support the fitness of this model to native, HCl and H<sub>2</sub>SO<sub>4</sub> modified biomasses in case of both sizes. However, values of  $R^2$ ,  $K_L$  and  $X_m$  (Table 1) for H<sub>3</sub>PO<sub>4</sub> modified sorbents indicated that Langmuir theory describes the sorption phenomena for this biomass more favorably.

#### 3.3.3. Temkin isotherm model

Temkin isotherm plot [26] was chosen to evaluate adsorption potentials of sorbent for sorbate. The heat of sorption of the molecules in layer decreases linearly with coverage due to sorbate and sorbent interactions [27]. Temkin isotherm assumes that the fall in the heat of sorption is linear rather than logarithmic. Temkin isotherm model is shown:

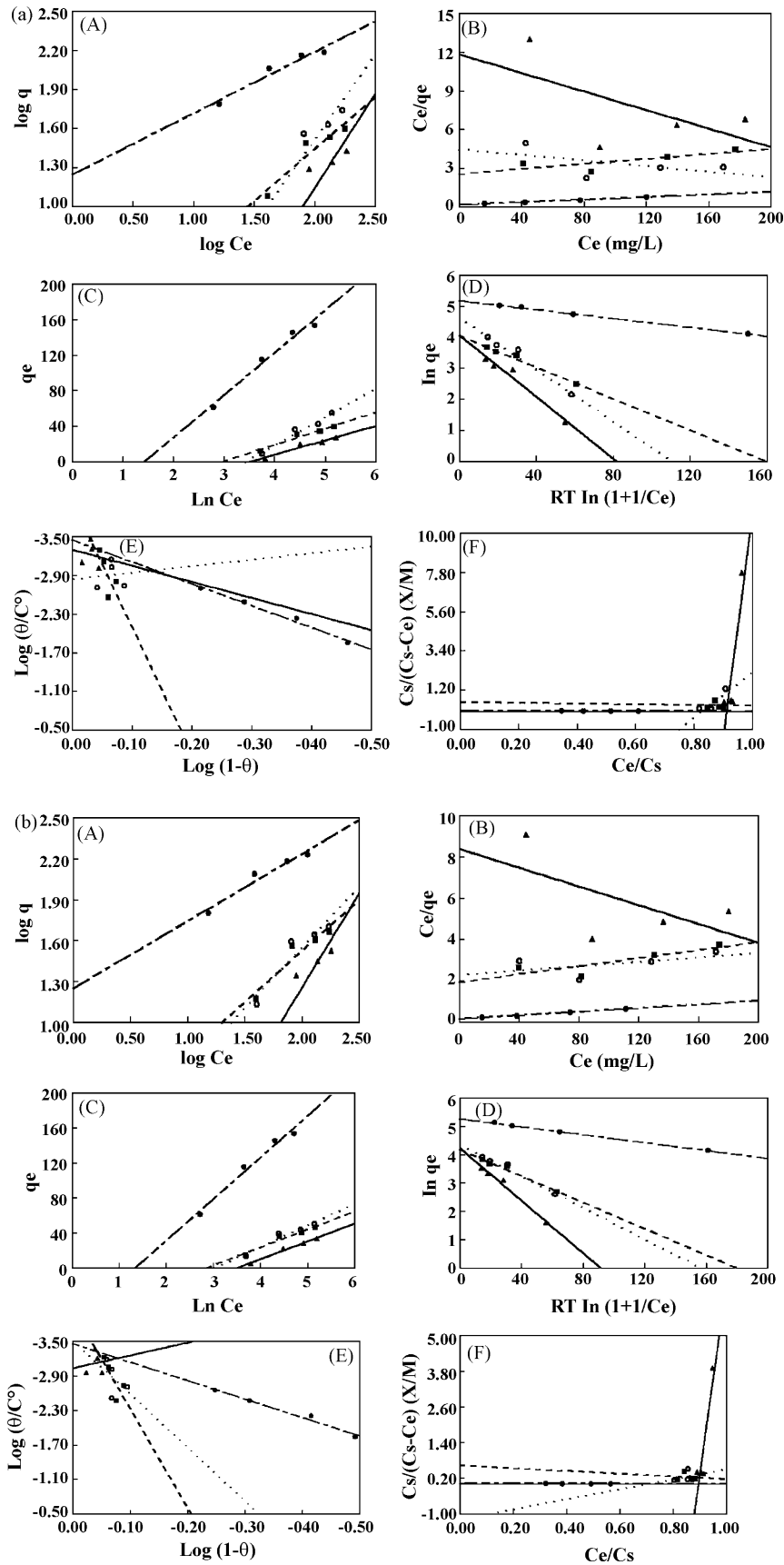
$$q_e = \frac{RT}{b_T} \ln K_T + \frac{RT}{b_T} \ln C_e \quad (4)$$

where  $K_T$  (dm<sup>3</sup>/g) is Temkin adsorption potential and  $b_T$  (kJ/mol) is heat of sorption (Table 1). When evaluated with respect to coefficient of determination ( $R^2$ ), Temkin isotherm model appears to be favorable for fitness to the experimental data. Values of  $K_T$  (dm<sup>3</sup>/g) for H<sub>3</sub>PO<sub>4</sub> modified biomass showed that model favors the experimental data of H<sub>3</sub>PO<sub>4</sub> modified biomass of both sizes. Order of  $b_T$  values was in accordance with the experimental findings because heat of sorption of the molecules decreases linearly with coverage due to sorbate and sorbent interactions and smaller values of  $b_T$  for H<sub>3</sub>PO<sub>4</sub> modified biomass concluded this biosorbent as more favorable.

#### 3.3.4. Dubinin–Radushkevich isotherm model

The Dubinin–Radushkevich model is used to estimate the characteristic porosity of the biomass and apparent energy of adsorption [28]. The model is represented by

$$\ln q_e = \ln q_D - 2B_D RT \ln \left( 1 + \frac{1}{C_e} \right) \quad (5)$$



**Fig. 3.** (a) Equilibrium isotherm analysis of  $Pb^{2+}$  biosorption using 63  $\mu m$  sieved *Cicer arietinum* biomass (control ▲, HCl ■,  $H_2SO_4$  ○ and  $H_3PO_4$  ●) by linear plots of (A) Freundlich (B) Langmuir (C) Temkin (D) Dubinin–Radushkevich (E) Florry–Huggins and (F) BET models. (b) Equilibrium isotherm analysis of  $Pb^{2+}$  biosorption using 150  $\mu m$  sieved *C. arietinum* biomass (control ▲, HCl ■,  $H_2SO_4$  ○ and  $H_3PO_4$  ●) by linear plots of (A) Freundlich (B) Langmuir (C) Temkin (D) Dubinin–Radushkevich (E) Florry–Huggins and (F) BET models.

**Table 1**  
Equilibrium isotherm model parameters for the biosorption of Pb (II) using native and acidically modified *Cicer arietinum* pod biomass.

Equilibrium models	Parameters	63 $\mu\text{m}$ sieved biomass				150 $\mu\text{m}$ sieved biomass			
		Native	HCl	H <sub>2</sub> SO <sub>4</sub>	H <sub>3</sub> PO <sub>4</sub>	Native	HCl	H <sub>2</sub> SO <sub>4</sub>	H <sub>3</sub> PO <sub>4</sub>
Freundlich	$1/n$	1.45	0.802	1.307	0.472	1.382	0.7524	0.8903	0.4956
	$k$ (dm <sup>3</sup> /g)	0.0179	0.6897	0.078	17.69	0.031	1.052	0.588	17.81
	$R^2$	0.9343	0.9515	0.9461	0.9711	0.955	0.9473	0.9390	0.9773
	$q_{\text{exp}}$ (mg/g)	26.9	39.7	55.08	153.8	33.62	46.3	50.4	171.28
Langmuir	$K_L$ (dm <sup>3</sup> /g)	$3.039 \times 10^{-3}$	$3.753 \times 10^{-3}$	$2.423 \times 10^{-3}$	$3.01 \times 10^{-2}$	$2.7 \times 10^{-3}$	$5 \times 10^{-3}$	$2.3 \times 10^{-3}$	$2.8 \times 10^{-2}$
	$X_m$ (mg/g)	27.789	103.69	92.16	200.36	44.01	102.93	191.31	228.52
	$R^2$	0.5894	0.7774	0.5211	0.9962	0.5937	0.8446	0.5238	0.9981
	$q_{\text{exp}}$ (mg/g)	26.9	39.7	55.08	153.8	33.62	46.3	50.4	171.28
Temkin	$b_T$ (kJ/mol)	155.75	137.16	78.798	52.745	123.96	122.59	101.92	53.05
	$K_T$ (dm <sup>3</sup> /g)	0.0299	0.0516	0.0326	0.2411	0.0299	0.0579	0.0482	0.263
	$R^2$	0.9749	0.9771	0.9816	0.9882	0.9931	0.9721	0.9699	0.9887
	$q_{\text{exp}}$ (mg/g)	26.9	39.7	55.08	153.8	33.62	46.3	50.4	171.28
Dubinin–Radushkevich	$B_D$ (mol <sup>2</sup> /kJ <sup>2</sup> )	0.2493	0.01273	0.02104	0.0359	0.0232	0.0116	0.0139	0.0351
	$q_D$ (mg/g)	58.52	58.89	106.52	179.97	66.523	68.96	79.96	196.42
	$R^2$	0.9808	0.9896	0.9850	0.9992	0.9917	0.9877	0.9845	0.9993
	$q_{\text{exp}}$ (mg/g)	26.9	39.7	55.08	153.8	33.62	46.3	50.4	171.28
Florry–Huggins	$n$	2.492	19.55	1.02	3.397	2.235	17.25	9.29	3.201
	$K_{\text{FH}}$	$5 \times 10^{-4}$	$8.4 \times 10^{-5}$	$1.44 \times 10^{-3}$	$3.5 \times 10^{-4}$	$9.2 \times 10^{-4}$	$8.4 \times 10^{-5}$	$2.9 \times 10^{-4}$	$3.4 \times 10^{-4}$
	$R^2$	0.1413	0.6924	0.0877	0.9936	0.1289	0.7654	0.4742	0.9889
	$q_{\text{exp}}$ (mg/g)	26.9	39.7	55.08	153.8	33.62	46.3	50.4	171.28
BET	$A$	1.105	1.553	1.3026	31.929	1.134	0.868	2.256	32.102
	$X_m$ (mg/g)	0.0079	1.2498	0.07262	1.028	0.0152	1.842	0.372	1.034
	$R^2$	0.8612	0.0225	0.8458	0.6279	0.8978	0.1047	0.2742	0.7179
	$q_{\text{exp}}$ (mg/g)	26.9	39.7	55.08	153.8	33.62	46.3	50.4	171.28

where  $q_D$  is Dubinin–Radushkevich isotherm constant related to the degree of sorbate sorption by sorbent surface and  $B_D$  is related to the free energy of sorption per mole of sorbate as it migrates to the surface of the biomass from infinite distance in the solution. The porosity parameter values 'B' for the biomass towards the metal ion were less than unity indicating that sorption of Pb (II) by native and pretreated biomass was significant (Table 1).  $q_D$  values obtained from isotherm data were in accordance with the experimental data of all sorbents.  $R^2$  values obtained from data also supported the fitness of model to all sorbents. Overall, this model explains the sorption phenomena for all biomasses.

### 3.3.5. Florry–Huggins isotherm model

The Florry–Huggins isotherm model [18] accounts the degree of surface coverage characteristic of the sorbate on the sorbent. The isotherm is as follows:

$$\log \frac{\theta}{C_0} = \log K_{\text{FH}} + n \log(1 - \theta) \quad (6)$$

where  $\theta = (1 - C_e/C_0)$  is degree of surface coverage,  $K_{\text{FH}}$  is equilibrium constant,  $n$  is metal ions occupying active sites of biomass,  $C_0$  is initial metal concentration and  $C_e$  is equilibrium metal concentration.

A greater value of 'n' means that greater number of metal ions are occupying the binding sites. Values of  $n$  (Table 1) obtained showed that results are not in good agreement with experimental  $q$  values. A greater value of 'n' in case of HCl modified biomass revealed that HCl modified biomass is more efficient than other biomasses for metal removal. Values of  $K_{\text{FH}}$  showed that H<sub>2</sub>SO<sub>4</sub> modified biomass is more favorable among 63  $\mu\text{m}$  sieved biomasses and native biomass is more favorable among 150  $\mu\text{m}$  sieved biomasses because larger value of  $K_{\text{FH}}$  indicates efficiency of biomass. These findings suggested that Florry–Huggins isotherm plot is not favorable to depict the sorption. However, values of correlation coefficients as 0.9936 in case of 63  $\mu\text{m}$  sieved and 0.9889 in case of 150  $\mu\text{m}$  sieved H<sub>3</sub>PO<sub>4</sub> modified biomasses revealed a linear relationship between

Florry–Huggins isotherm model and H<sub>3</sub>PO<sub>4</sub> modified biomass as compared to other biomasses.

### 3.3.6. BET isotherm model

The BET model assumes that a number of layers of adsorbate molecules are formed at the surface of sorbent and the Langmuir equation applies to each layer [29]. A further assumption of the BET model is that a given layer need not complete formation prior to initiation of subsequent layers; the equilibrium condition will therefore involve several types of surfaces in the sense of number of layers of molecules on each surface. For adsorption from solution with the additional assumption that layers beyond the first have equal energies of adsorption, the BET equation takes the simplified form as

$$\frac{C_s}{(C_s - C_e)(X/M)} = \frac{1}{AX_m} + \frac{(A - 1)}{AX_m} \left\{ \frac{C_e}{C_s} \right\} \quad (7)$$

where  $(X/M)$  is the amount of solute adsorbed per unit weight of the adsorbent (mg/g),  $C_e$  is the equilibrium concentration of the solute in bulk solution (mg/L),  $C_s$  is the saturation concentration of the solute in bulk solution (mg/L),  $A$  is a constant indicative of the energy of interaction between the solute and the adsorbent surface and  $X_m$  is a constant indicative of the amount of solute adsorbed forming a complete monolayer (mg/g). Values of  $R^2$  indicated BET model not appropriate to describe the sorption mechanism (Table 1) for both sizes of native and pretreated biomasses. Also, there is much difference between values of  $X_m$  and  $q_{\text{exp}}$ .

### 3.3.7. Correlation coefficient

The correlation coefficients ' $R^2$ ' determined from linearization of the six two-parameter isotherms (Table 1) suggest that Temkin and Dubinin–Radushkevich isotherms are most appropriate to explain the sorption phenomena in case of all biomasses whereas Freundlich and Florry–Huggins isotherms describe sorption phenomena in case of H<sub>3</sub>PO<sub>4</sub> modified biomass also. To solve this complication, statistical analysis (two-way ANOVA) at  $\alpha = 0.05$  were

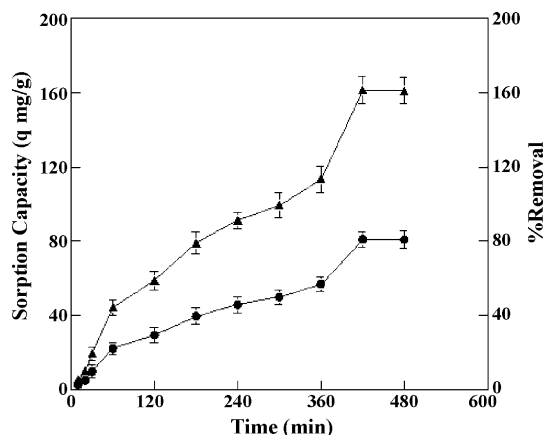
**Table 2**Two-way analysis of variance (ANOVA) without replication at  $\alpha = 0.05$  to evaluate the relationship between isotherm models and biomass.

	Count	Sum	Average	Variance	
<b>a</b>					
Freundlich	8	7.6216	0.9527	0.000221346	
Langmuir	8	5.8443	0.7305	0.04037448	
Temkin	8	7.8456	0.9807	$7.31457 \times 10^{-5}$	
Dubinin–Radushkevich	8	7.9178	0.9897	$4.55364 \times 10^{-5}$	
Florry–Huggins	8	4.2724	0.534	0.145411671	
BET	8	4.352	0.544	0.127413246	
Blank (63 $\mu\text{m}$ )	6	4.4819	0.7469	0.109286806	
HCl (63 $\mu\text{m}$ )	6	4.4105	0.7351	0.13631491	
H <sub>2</sub> SO <sub>4</sub> (63 $\mu\text{m}$ )	6	4.3673	0.7279	0.128919606	
H <sub>3</sub> PO <sub>4</sub> (63 $\mu\text{m}$ )	6	5.5762	0.9294	0.021910899	
Blank (150 $\mu\text{m}$ )	6	4.5602	0.7600	0.118195447	
HCl (150 $\mu\text{m}$ )	6	4.6218	0.7703	0.113576572	
H <sub>2</sub> SO <sub>4</sub> (150 $\mu\text{m}$ )	6	4.1656	0.6943	0.094809551	
H <sub>3</sub> PO <sub>4</sub> (150 $\mu\text{m}$ )	6	5.6702	0.9450	0.012444539	
Source of variance	Degree of freedom	Sum of squares	Mean sum of squares	$F_{\text{cal}}$	$F_{\text{tab}}$
<b>b</b>					
Biomass	7	0.375713135	0.053673305	1.032711	2.28523517
Isotherm	5	1.858228799	0.37164576	7.150716	2.48514322
Error	35	1.819062839	0.051973224		
Total	47	4.053004773			

done to know about model fitted mostly to experimental data. This method describes the relationship between the isotherm models in describing the sorption systems and between the biomasses for metal ion removal. The summary of the statistical analysis is presented in Table 2a and b. By considering Table 2a, it is clear that Dubinin–Radushkevich isotherm model provides a better fit for the sorption systems because of larger values of sum, average and smaller values of variance of correlation coefficients. Also, it is clear in case of H<sub>3</sub>PO<sub>4</sub> modified biomass that sum and average of correlation coefficients for this biomass are large but variance is small (Table 2a). So, H<sub>3</sub>PO<sub>4</sub> modified biomass showed remarkable increase in sorption capacity. However, the results of ANOVA (Table 2b) indicate that there is no significant difference among the biomasses for sorption. This indicates that all biomasses can remove metal ions from aqueous solutions. Experimental results also showed that Dubinin–Radushkevich isotherm model is more favorably suitable to explain the sorption phenomena.

### 3.4. Kinetic study

The kinetic profiles of Pb (II) biosorption by *C. arintinum* biomass are shown in Fig. 4. The biosorption of Pb (II) onto *C. arintinum* biomass was very slow with 6.269% removal within first



**Fig. 4.** Effect of time interval on the biosorption of Pb<sup>2+</sup> (sorption capacity  $q$ , ▲ and % removal, ●) by H<sub>3</sub>PO<sub>4</sub> modified biomass.

20 min. Equilibrium was attained within 7 h. It is well known fact that rapid initial sorption is due to extracellular binding and slow sorption results from intracellular binding [30–32]. In this study, slow sorption may be due to intracellular binding. Saeed et al. [33] studied removal and recovery of lead (II) from single and multi-metal (Cd, Cu, Ni, Zn) solutions by black gram husk and reported that sorption rate was very rapid and equilibrium was attained within 30 min. In this study, pretreatment of biomass by H<sub>3</sub>PO<sub>4</sub> leads to slow sorption. Although pretreatment caused changes in active sites of biomass that led to slow sorption but pretreatment enhanced sorption capacity of biomass. Shreedhara and Ryan [34] investigated mercury, cadmium, copper, lead and uranium adsorption on cellulose–dithiocarbamate resins and reported that the adsorption rate was very slow. Actually, there are several parameters which affect biosorption rate. These parameters include stirring rate of aqueous phase, the amount of sorbent, the structural properties of biosorbent, the presence of co-ions and the properties of ion under study. Therefore, it is possible that biosorbents vary in their removal rates of metal ions.

### 3.5. Kinetic modeling

Kinetics of metal sorption governs the rate that determines the residence time and it is one of the important characteristics defining the efficiency of an adsorbent. Sorption kinetics can be controlled by several independent processes that could act in parallel or in series such as external mass transfer (film diffusion), bulk diffusion, intraparticle diffusion [35] and chemisorption (chemical reaction). Film transport is diffusion of heavy metals through boundary around biosorbent [36] whereas bulk diffusion is transport of metal ions in solution [37]. Chemisorption is chemical reaction between active sites of biomass and heavy metal ions [36] and intraparticle diffusion is diffusion of metals from surface to internal sites [38]. Kinetic equations have been developed to explain the transport of metals onto various adsorbents (Fig. 5). These equations include the pseudo-first-order equation [39], the pseudo-second-order equation [40], the elovich equation [38,41], intraparticle diffusion model [42], saturation (mixed order) model [43], second-order rate equation model [43] and bangham model [44]. The pseudo-first-order is given by

$$\log(q_e - q) = \log q_e - \frac{k_{1,\text{ads}}t}{2.303} \quad (8)$$



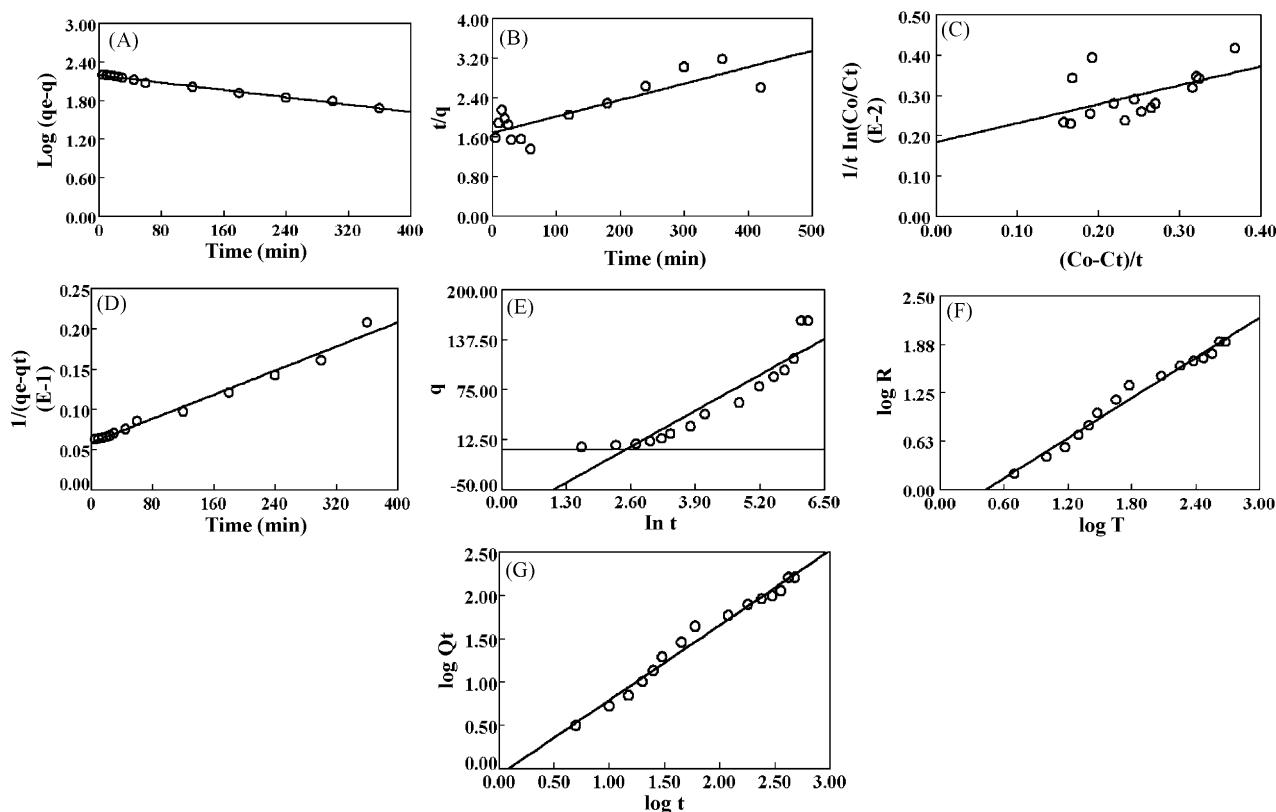


Fig. 5. Kinetic study of  $\text{Pb}^{2+}$  biosorption by linear plots of (A) pseudo-first-order (B) pseudo-second-order (C) saturation (D) second-order rate equation (E) Elovich (F) intraparticle diffusion and (G) Bangham models using  $\text{H}_3\text{PO}_4$  modified biomass.

where  $q_e$  (mg/g) and  $q$  (mg/g) are the adsorption capacities at equilibrium and at time  $t$ , respectively,  $k_{1,\text{ads}}$  is the rate constant of pseudo-first-order adsorption ( $\text{min}^{-1}$ ).  $k_{1,\text{ads}}$  and  $q_e$  can be determined from slope and intercept, respectively.

The pseudo-second-order reaction kinetic is expressed as

$$\frac{t}{q} = \frac{1}{k_{2,\text{ads}}q_e^2} + \frac{t}{q_e} \quad (9)$$

where  $k_{2,\text{ads}}$  is the rate constant of pseudo-second-order adsorption ( $\text{g}/(\text{mg min})$ ) and  $q_e$  is the adsorption capacity at equilibrium (mg/g).

The saturation (mixed order) model is given as

$$\frac{1}{t} \ln \frac{C_0}{C_t} = -\frac{k_0}{K} - \frac{1}{K} \left( \frac{C_0 - C_t}{t} \right) \quad (10)$$

where  $C_0$  and  $C_t$  are the concentrations of metal ion (mg/L) at  $t=0$  and at any time, respectively.  $k_0$  ( $\text{L}/(\text{g min})$ ) and  $K$  ( $\text{L}/\text{mg}$ ) are saturation rate constants of adsorption.

Second-order rate equation is expressed as

$$\frac{1}{(q_e - q_t)} = \frac{1}{q_e} + k^0 t \quad (11)$$

where  $q_e$  and  $q_t$  are the amounts of metal ion sorbed at equilibrium and at any time (mg/g), respectively whereas  $k^0$  is second-order rate constant of adsorption ( $\text{g}/(\text{mg min})$ ).

Bangham model is given as

$$\log Q_t = \log K_r + \left( \frac{1}{m} \right) \log t \quad (12)$$

where  $K_r$  (mg/(g min)) and  $1/m$  are the sorption rate constants. A larger value of  $K_r$  depicts the suitability of biosorbent for metal removal and  $1/m$  tells that kinetic was applicable to sorption phenomena up to this extent.

Elovich model is described as

$$q_t = \frac{1}{\beta} \ln(\alpha\beta) + \frac{1}{\beta} \ln(t) \quad (13)$$

where  $\alpha$  is initial adsorption rate (mg/(g min)),  $\beta$  is the desorption constant (g/mg).

The intraparticle diffusion model proposed by Webber and Morris [42] is expressed as

$$\log R = \log K_{\text{id}} + a(\log t) \quad (14)$$

where  $R$  is % removal,  $t$  is contact time,  $a$  is gradient of linear plot and it depicts the adsorption mechanism,  $K_{\text{id}}$  is intraparticle diffusion rate constant ( $\text{min}^{-1}$ ) and can be written as rate factor which describes about percent heavy metal adsorbed per unit time. Comparison of various kinetic model parameters is tabulated in Table 3. The Lagergren pseudo-first-order rate constant ( $k_{1,\text{ads}}$ ) and  $q_e$  determined from model along with correlation coefficient indicated that  $q_e$  is very close to  $q_{\text{exp}}$  and correlation coefficient is also greater than 0.98. It suggests that adsorption was a first-order reaction. Pseudo-second-order rate constant ( $k_{2,\text{ads}}$ ) and  $q_e$  values determined from pseudo-second-order model suggested that sorption rate cannot be explained by pseudo-second-order reaction.  $q_e$  differs substantially from those measured experimentally and correlation coefficient is also less than measured in case of pseudo-first-order model. Value of  $k_{2,\text{ads}}$  is less than  $k_{1,\text{ads}}$  suggesting that pseudo-first-order reaction can explain sorption rate. The second-order rate equation described the biosorption kinetic of Pb (II) with high correlation coefficient ( $R^2 = 0.9913$ ) and  $q_e$  was also close to  $q_{\text{exp}}$  whereas saturation and elovich models were unable to explain the sorption rate because of correlation coefficient ( $R^2 < 0.98$ ). In elovich model,  $\alpha$  is related to rate of chemisorption (initial adsorption rate) and  $\beta$  is related to surface coverage (desorption constant). Elovich model describes about heterogeneous sorption and a small value of  $\beta$  tells

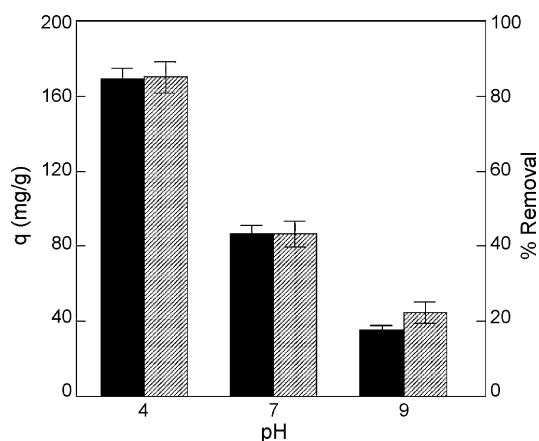
**Table 3**  
Kinetic model parameters for the biosorption of Pb (II) using H<sub>3</sub>PO<sub>4</sub> modified *C. arientinum* pod biomass.

Kinetic models	Kinetic parameter values		
Pseudo-first-order model	$k_{1,ads}$ (min <sup>-1</sup> ), $3.298 \times 10^{-3}$	$q_e$ (mg/g), 156.91	$R^2$ , 0.9948
Pseudo-second-order model	$k_{2,ads}$ (g/(mg min)), $6.569 \times 10^{-6}$	$q_e$ (mg/g), 300.66	$R^2$ , 0.8507
Saturation model	$K$ (L/mg), 212.31	$k_0$ (L/(g min)), 0.3911	$R^2$ , 0.5275
Second-order rate equation model	$k^0$ (g/(mg min)), $4 \times 10^{-5}$	$q_e$ (mg/g), 171.09	$R^2$ , 0.9913
Bangham's model	$K_f$ (mg/(g min)), 0.8401	$m$ , 1.156	$R^2$ , 0.9905
Elovich model	$\alpha$ (mg/(g min)), 2.836	$\beta$ , 0.0289	$R^2$ , 0.9324
Intraparticle model	$a$ , 0.8652	$K_{id}$ (min <sup>-1</sup> ), 0.4212	$R^2$ , 0.9905
			$q_{exp}$ (mg/g), 161.42

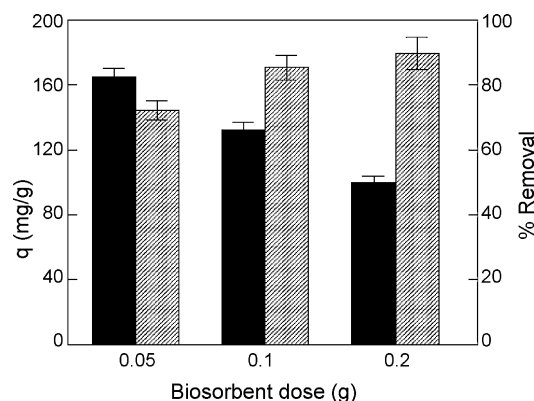
that model is unable to explain heterogeneous surface of biomass. A slow sorption rate and small value of constant  $\alpha$  tells that no chemisorption reaction occurred. Hence this model cannot provide an accurate fit to experimental data. Bangham assumption tells that biosorption of Pb (II) followed the (1/ $m$ )th order sorption rate and constant  $K_f$  was 0.8401 (mg/(g min)). Although, value of  $K_f$  is small but value of  $R^2$  tells that mechanism told by Bangham was also involved during description of sorption rate. Value of  $a$  gives an idea about the thickness of boundary layer, i.e., the larger the intercept ( $a$ ) value, the greater will be the boundary layer effect [45].  $K_{id}$  is rate factor i.e., percent heavy metal adsorbed per unit time and its value tells that percent heavy metal adsorbed was 0.4212 per unit time. The linearity of this plot ( $R^2 = 0.9905$ ) describes the involvement of intraparticle diffusion during this study up to some extent. Since several independent processes can describe kinetics, therefore applicability of more than one independent sorption rate mechanisms is not a problem.

### 3.6. Effect of pH

It is well known that sorption of heavy metal ions by solid substrates depends on the pH of the solution. Fig. 6. shows that uptake capacity and % removal of Pb (II) by *C. arientinum* decreased with increase in pH. It is generally known that at low pH values of 1–3, solution is strongly acidic and the surface of the sorbent is surrounded by hydrogen ions, which prevent the metal ions from approaching the binding sites on the sorbent. Due to this reason, it is thought that sorption capacity increases with increase in pH. But it is important that during this study, minimum pH for solution media was 4 and a pH of 4 is not strongly acidic but slightly acidic. At pH value of 4, metal ions can easily compete hydrogen ions and bind active sites of biosorbent. Due to this reason, sorption capacity and % removal was high at pH 4. At pH 7 and 9, sorption capacity and % removal decreased and this decrease can be explained by the fact that at higher pH, number of hydroxyl ions is more and chances of formation of metal hydroxides are also more that result in pre-



**Fig. 6.** Effect of pH on the biosorption of Pb<sup>2+</sup> ( $q$  values: ■, % removal: ▨) by H<sub>3</sub>PO<sub>4</sub> modified biomass.



**Fig. 7.** Effect of biosorbent dose on the biosorption of Pb<sup>2+</sup> ( $q$  values: ■, % removal: ▨) by H<sub>3</sub>PO<sub>4</sub> modified biomass.

cipitation. This precipitation caused a decrease in sorption capacity and % removal with increase in pH.

### 3.7. Effect of biosorbent dose

Biosorption is dependent largely on biosorbent dose. Effect of biosorbent dose was studied using H<sub>3</sub>PO<sub>4</sub> pretreated *C. arientinum* biomass having 150  $\mu$ m sieved particle size added to 100 mL of 100 mg/L of Pb<sup>2+</sup>. A biosorbent dose of 0.05, 0.1 and 0.2 g having mentioned particle size was used to evaluate the effect of biosorbent dose. Fig. 7 shows that with increase in biosorbent dose, sorption capacity decreases but percentage removal increases. A similar behavior was shown by work done by Abu Al-Rab et al. [46]. They observed that during work on biosorption of copper on *Chlorella vulgaris* from single, binary and ternary metal aqueous solutions, sorption capacity decreases but % removal increases with increase in biosorbent dose. The decrease in metal uptake by increasing sorbent dose may be due to many reasons, such as, electrostatic interactions, interference between binding sites, availability of solute and reduced mixing at high biomass densities [47,48]. It is concluded that because of lower adsorptive capacity of the sorbent, the adsorbent active sites remain unsaturated during the sorption process and due to this, sorption efficiency decreases. Some of these reasons also contribute in limiting the maximum percentage removal and due to this 100% removal was not attained.

## 4. Conclusion

This study investigated the use of acidically modified *C. arientinum* biomass for the removal of lead (II) from aqueous solutions. Pretreatment enhanced the biosorption capacity of biomass and H<sub>3</sub>PO<sub>4</sub> treated biomass showed maximum sorption capacity. The sorption capacity increased in the order H<sub>3</sub>PO<sub>4</sub> > H<sub>2</sub>SO<sub>4</sub> > HCl > native, with increase in metal concentration and % removal was maximum at 100 mg/L and then decreased. Sorption rate was very slow and equilibrium was attained within 7 h. In case of equilibrium study, Dubinin–Radushkevich isotherm



plot was found in better correlation with experimental data. Sorption rate was best explained by pseudo-first-order, second-order rate equation, bangham and intraparticle diffusion models because sorption kinetics can be described by several independent processes that could act in parallel or in series. When evaluated the effect of biosorbent dose, sorption capacity decreased but % removal increased with increase in biosorbent dose. At pH 4, maximum sorption capacity (169.23 mg/g) was attained.

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